



PATIENT

Lucy Smith

SPECIES

Feline

BREED

Domestic Shorthair

SEX

Spayed female

AGE

9 years

WEIGHT

11.7 lbs

INTERPRETED BY

Eric Lindquist, DMV
DABVP, Cert. IVUSS

IMAGING PERFORMED BY

Ian Anderson

HOSPITAL NAME

Chester AC

REFERRING VET

Dr. Angela Feiring

INVOICE

78413

DATE

6/8/26

PRESENTING CLINICAL SIGNS

History: Two week history of intermittent vomiting.
Moderate hypercalcemia (confirmed with ionized calcium)

ULTRASONOGRAPHIC EXAMINATION OF THE ABDOMEN

Urinary System

The **urinary bladder**, trigone, and pelvic urethra presented normal thicknesses and normal tone. The ureters were not visible which is normal. No uroliths or sediment were visualized and anechoic urine was present. No evidence of inflammatory or neoplastic changes was noted. Ureteral papillae were normal.

The **kidneys** revealed largely normal size and structure, corticomedullary definition and ratio (cortex 1/3 of medulla) were essentially maintained with some age-related loss of curvilinear patterns regarding the capsule and C/M junction. The cortices presented largely uniform texture with some increased echogenicity expected for this age patient. Medullary structure differed distinctly from that of the cortex and no evidence of pelvic dilation was present. Slight pinpoint mineralization was noted. Slight cortical infarct is noted at the cranial pole of the left kidney. The patient likely has a history of moving calculi. Blood flow appeared adequate on the left kidney. The left kidney measured 3.9 cm. The right kidney measured 3.9 cm.

Adrenal Glands

Both **adrenal glands** were visualized and recognized as having normal shape, size, position and echogenicity for this breed. The phrenic vasculature, glandular echogenicity and detail were unremarkable. Capsule, cortex, and medullary definition were normal for this age patient.

Spleen

The **spleen** presented a smooth homogeneous parenchyma hyperechoic to liver and renal cortical parenchyma. The capsule was smooth without noticeable expansion or deviation from within the spleen or adjacent pathology. The splenic vasculature demonstrated normal volume without signs of congestion or thrombosis. No sonographic evidence of acute or chronic inflammatory, neoplastic, or infarctual changes was noted. The spleen measured 0.9 cm.

Liver

The **liver** images submitted revealed subjectively normal liver size, contour, and structure. Parenchymal echogenicity was naturally coarse and hypoechoic to the spleen. Vascular and biliary tracts were of normal volume with no evidence of congestion. The gallbladder presented acceptably thin walls with primarily anechoic content. The cystic and common bile ducts were normal. No pathological hepatic lymphadenopathy was evident. No overt structural evidence of inflammatory, infiltrative or regenerative pathology was evident.



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Gastrointestinal

Examination of the **gastrointestinal tract** revealed a stomach and intestine free of stasis, of normal wall thickness, acceptable curvilinear mural detail, and peristaltic activity. Small and large intestine demonstrated normal luminal chyme and stool consistency respectively. No obstructive or overt infiltrative disease was noted. No associated abnormal lymphatic activity was noted.

Pancreas

The base and limbs of the **pancreas** were observed to be largely isoechoic to surrounding omental fat. Pancreatic duct and capsular contour were acceptably normal and parenchyma respected normal curvilinear patterns. No overt evidence of active inflammatory or neoplastic disease was noted.

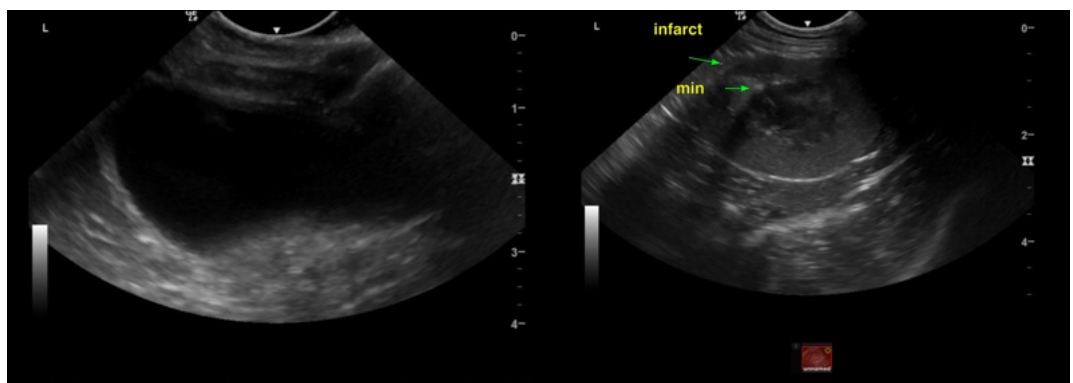
ULTRASONOGRAPHIC FINDINGS

Structurally unremarkable abdomen

Minor pinpoint nephrolithiasis.

INTERPRETATION OF THE FINDINGS & FURTHER RECOMMENDATIONS

There was no evidence of primary disease responsible for the hypercalcemia. Full hypercalcemia panel is warranted. Chest radiographs, spinal radiographs are recommended to assess for lytic disease +/- parathyroid imaging/ultrasound.





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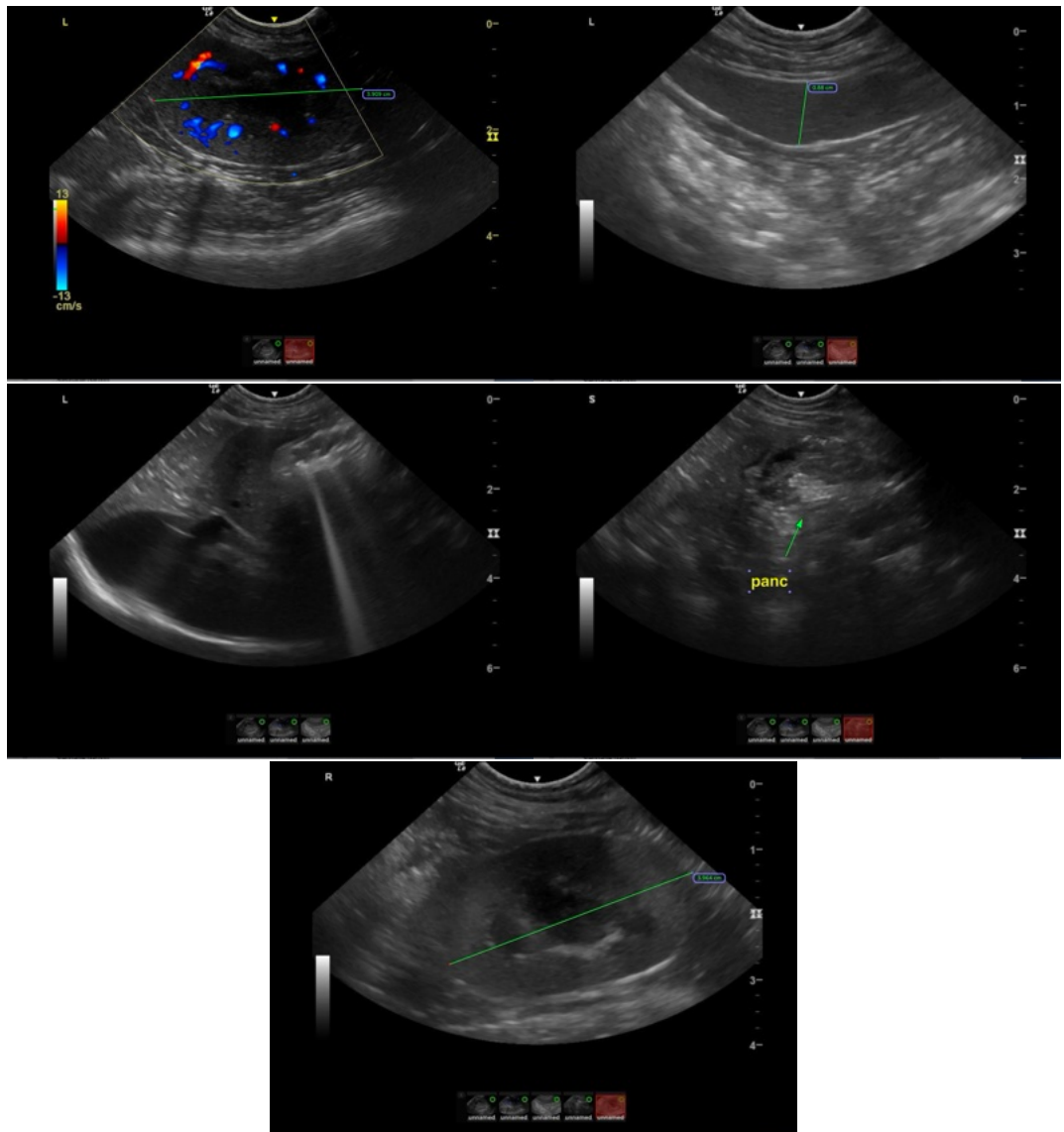
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The information and recommendations provided are based on the images presented by the referring veterinarian/sonographer. No evaluation can be communicated regarding pathology that was not visible in the image/video clips provided.

Thank you for this referral. If the clinical or image interpretation does not parallel your findings or if I can be of any further assistance please contact me.

Eric Lindquist, DMV, DABVP (CFM), Cert. IVUSS, CEO of SonoPath.com

info@SonoPath.com



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Excerpt from the Curbside Guide: <https://sonopath.com/thecurbsideguide/>

Hypercalcemia (Feline)

DESCRIPTION Feline hypercalcemia can be a frustrating diagnosis for practitioner and concerned cat owner alike. When making a diagnosis of hypercalcemia, it is important to consider the physiology of calcium homeostasis. Calcium levels measured using standard chemistry profiles represent the total serum calcium, which is comprised of 1) calcium bound to proteins complex with anions, such as citrate and phosphate, and 2) ionized or “free” calcium, which is the metabolically active form of the element. The total calcium level should not, however, be used to make a diagnosis of hypercalcemia, as it is influenced by the patient’s acid-base status and by various bound proteins, specifically albumin. To this end, the formula used to correct for calcium is based on albumin levels. Only the ionized calcium level can be utilized to arrive at an accurate diagnosis of hypercalcemia.

CAUSES OF HYPERCALCEMIA Hyperparathyroidism Benign adenomas of the parathyroid gland create an excess of parathyroid hormone (PTH), which in turn stimulates calcium release from bones and increases renal reabsorption of calcium while promoting phosphorus excretion. A diagnosis can be made by evaluating a paired PTH/ionized calcium sample. Generally, high PTH combined with elevated ionized calcium is indicative of primary hyperparathyroidism; however, one may find that even when ionized calcium is high, PTH levels may still fall within the “normal” range. An inappropriately “high-normal” PTH level, in conjunction with an elevated ionized calcium level, may therefore indicate primary hyperparathyroidism, especially if it is clinically consistent with the signalment (primarily geriatric patients) and clinical findings. In such cases, reevaluation of the panel in six weeks is recommended to determine whether the PTH is trending upwards. Sonographic examination of the parathyroid glands can also be performed, as hypertrophy of a single parathyroid or an associated nodule may be discovered. It is common for patients with hyperparathyroidism to also display hypophosphatemia, which may aid in the diagnosis of borderline cases.

1. Renal Disease It has long been thought that renal failure is a common cause of hypercalcemia, primarily because it increases calcium complexes, which in turn lead to elevated total serum calcium. Yet, ionized calcium frequently remains at normal levels during renal failure, and patients in these circumstances are not truly hypercalcemic. Some cats with renal compromise do display both elevated ionized calcium and PTH (tertiary hyperparathyroidism). Although the mechanism is unclear, it is thought that parathyroid glands affected by chronically elevated calcium levels become autonomous (i.e., they no longer respond to calcium levels). In fact, the elevated level of calcium becomes dependent on excess PTH instead of PTH rising in response to low blood calcium levels. There has been evidence of this in human renal transplant patients who continue to be hypercalcemic after successful transplantation, and this is corroborated further by sonographic evaluations that reveal global hypertrophy of the parathyroid glands. These patients also tend to become azotemic and hyperphosphatemic, which is consistent with renal compromise.

2. Hypercalcemia of malignancy A variety of neoplastic conditions are related to hypercalcemia: in cats, the most common are lymphoma and squamous cell carcinoma of the head and neck. Other causes include carcinomas originating in other tissues, multiple myeloma, leukemia, and sarcomas (e.g., osteosarcoma and fibrosarcoma). Hypercalcemia of malignancy is caused primarily by systemic humoral factor stimulation, which further incites osteoclastic activity. The most important factor is a fetal protein called PTH-related protein (PTHrP). This protein acts like PTH and can be detected in the serum. It should be noted that although a positive result indicates hypercalcemia of malignancy, a negative result does not rule it out. Moreover, hematopoietic bone neoplasms, such as multiple myeloma or leukemia, or primary or metastatic bone tumors, can cause local osteolytic effects. In multiple myeloma, plasma cells also secrete an osteoclastic activation factor. Further diagnostics should include, at a minimum, a thorough physical examination, abdominal ultrasound, and thoracic radiographs. A tissue biopsy or aspirate or a bone marrow evaluation may also be necessary. Skeletal radiographs or bone scans may also be required to help diagnose bone or hematopoietic neoplasia. Interestingly, hypercalcemia related to lymphoma often resolves abruptly with chemotherapy. Hypercalcemia related to carcinoma (if inoperable) and other neoplasms may be managed by glucocorticoid therapy and fluid administration as well as bisphosphonates.



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3. Idiopathic hypercalcemia Also called “benign hypercalcemia”, idiopathic hypercalcemia frustrates veterinarians and cat owners alike. It refers to a condition where the total calcium is mildly to moderately elevated (usually < 15 mg/dL) and ionized calcium is high, yet no cause can be identified. PTH is normal or low, and PTHrP is not detectable. Some patients display no obvious clinical signs, while others present with nonspecific clinical signs, including weight loss, anorexia, vomiting, and constipation. In addition, signs associated with calcium urolithiasis (i.e., dysuria, stranguria, hematuria, abdominal pain) may exist as these patients are predisposed secondarily to increased calciuresis. Note that this diagnosis can be made only after all other potential causes of hypercalcemia have been ruled out.

4. Less common causes of hypercalcemia Vitamin D toxicity needs to be ruled out and a thorough evaluation of the patient’s history should be obtained to determine whether accidental ingestion might have occurred. Vitamin D levels can also be evaluated in the serum. Granulomatous diseases, particularly fungal infections, can also lead to a hypercalcemic state and osteolytic lesions depending on the site of infection. Elevated globulin levels may be seen, and fungal serology should be evaluated.

TREATMENT Treatment is indicated if calcium is chronically elevated (> 13 mg/ dL), patients display clinical signs of calcium urolithiasis, or there are signs of renal failure. Patients in renal failure require diuresis and should undergo monitoring of their calcium blood level. It should be noted that diuretics that increase calciuresis may incite urolithiasis and should be avoided. Other effective treatments that may help include both diet change and pharmaceutical intervention. Typically, alkalinizing diets are lower in calcium and phosphorus, help raise the pH—which may correct

hypercalcemia—and decrease calciuresis, thereby diminishing the likelihood of calcium oxalate stone formation. High-fiber diets may help decrease calcium absorption from the intestine; however, the nutrient content should be evaluated thoroughly as some

of these diets may contain elevated levels of calcium and phosphorus. Renal diets are less acidifying and lower in calcium and phosphorus; however, they are also lower in protein, which is not ideal for most cats, and diminished phosphorus may stimulate calcitriol production. Homemade calcium-restricted diets and low-carbohydrate/high-protein diets may also be helpful in managing hypercalcemia. Ideally, wet food should be administered as it helps to reduce urine concentration and diminish the formation of calcium-based uroliths.

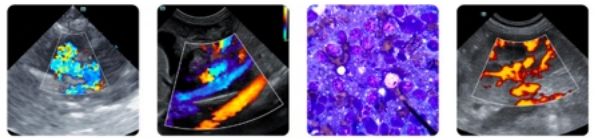
Glucocorticoids aid in decreasing intestinal calcium absorption but may increase renal excretion; monitoring the development of uroliths is therefore indicated. Glucocorticoids have been shown to decrease ionized calcium levels, yet high dosages appear to be necessary (1–2 mg/kg/day). In general, the owner should be prepared to repeat diagnostics (i.e., blood work, ultrasound, radiographs) to evaluate for emerging neoplasia, especially if the patient displays continued or worsening clinical signs. If the patient is nonresponsive, then alendronate (Fosamax) should be given at 10 mg/cat once a week and can be up-titrated to 30 mg/cat/week if necessary. Bisphosphonates act by inhibiting osteoclastic bone reabsorption through the obstruction of enzymatic pathways. These have been shown to be useful in both reducing hypercalcemia and potentially inhibiting tumor angiogenesis. (Note: Oral bisphosphonates can result in esophageal damage and strictures in people.) Give 5–6 mL of water following the administration of a pill as a preventative; alternatively, injectable forms can be utilized.

One can follow an algorithm-based approach to diagnosing hypercalcemic cats based on serum phosphorus levels:

gPO₄ i **Renal failure** i evaluate renal enzymes, urinalysis, blood pressure

Vitamin D toxicity i evaluate serum vitamin D concentration and history

Osteolytic disease i thorough physical examination, skeletal radiographs, +/- fungal serology



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kPO4 **i** **Hyperparathyroidism** **i** evaluate PTH level and ionized calcium, ultrasound of parathyroid glands

Malignancy **i** thorough physical examination, abdominal ultrasound, thoracic radiographs, PTHrP, and ionized calcium +/- tissue biopsy/ aspirate or bone marrow evaluation

REFERENCES

1. Audrey K and Cook AK. Cats & calcium: the causes & consequences of hypercalcemia. Proceedings from the American College of Veterinary Internal Medicine and Canadian Veterinary Medical Association, Montreal, QC, June 3–6, 2009.
2. Chew DJ, Schenck PA, Jaege JQ. Clinical disorders of hypercalcemia and hypocalcemia in dogs and cats.
3. de Brito Galvao JF, Chew DJ. Management of idiopathic hypercalcemia In: Little SE, ed. August's Consultations in Feline Internal Medicine. 7th ed. St. Louis, MO: Elsevier; 2013.
4. Hostutler RA, Chew DJ, Jaeger JQ, et al. Uses and effectiveness of pamidronate disodium for treatment of dogs and cats with hypercalcemia. *J Vet Intern Med* 2005; 19 (1): 29-33.
5. Middliff A, Chew D, Randolph J, et al. Idiopathic hypercalcemia in cats. *J Vet Intern Med* 2000; 6: 619-26.
6. Peterson ME. Hypercalcemia in dogs & cats: differential diagnosis & treatment. Proceedings from the Western Veterinary Conference, Las Vegas, NV, February 19–23, 2012.
7. Peterson ME. Pitfalls in the management of idiopathic hypercalcemia in cats. Proceedings from the American College of Veterinary Internal Medicine, Seattle, WA, June 11–15, 2013.
8. Schenck PA, Chew DJ. Investigation of hypercalcaemia and hypocalcaemia In: Mooney CT, Peterson ME, eds. BSAVA Manual of Canine and Feline Endocrinology. 4th ed. Quedgeley, Gloucester, UK: British Small Animal Veterinary Association; 2012: 221-33.
9. Whitney JL, Barrs VR, Wilkinson MR, et al. Use of bisphosphonates to treat severe idiopathic hypercalcaemia in a young Ragdoll cat. *J Feline Med Surg* 2011; 13 (2): 129-34.