



**PATIENT**

Emma McPherson

**SPECIES**

Canine

**BREED**

Yorshire Terrier

**SEX**

Spayed Female

**AGE**

6 Years

**WEIGHT**

5.1 Lbs.

**INTERPRETED BY**

Eric Lindquist, DMV  
DABVP, Cert. IVUSS

**IMAGING PERFORMED BY**

Dr. Erica Harmon

**HOSPITAL NAME**

Willamette VH

**REFERRING VET**

Dr. Erica Harmon

**INVOICE**

13146

**DATE**

12/25/21

**PRESENTING CLINICAL SIGNS**

History: P presented to rDVM 12/20 for 2 week history of respiratory concerns, o declined diagnostics at that time, p was painful on abdominal palpation, Convenia and ondansetron administered injectably and drontal dispensed. P rechecked 12/22 for recommended labwork and further care, once BW performed referral for continued care recommended, p presented 12/22-declined further diagnostics, outpatient care provided, SQ fluids, RX cerenia and rec OTC tums for Ca supplementation. P developed muscle spasms in hind limbs, diarrhea and some vomiting and represented 12/24, free abdominal fluid noted, abdominal pain, palpable fluid wave, dehydration

Abnormal PE/Chem/CBC/UA Results: Labs: 12/20 BW- rDVM transfer from Paws- Calc 4.3, TP 3.1, Alb 1.2, Glob 1.9, abnormal cPL, WBCs 20.8k, 12/22 PM bloodwork- EPOC- CA 0.71, LAC 3.66, HCT 48% 12/24 AM bloodwork- CBC- Hct wnl 42.8%, Retic 9.1 K/uL, mild-mod leukocytosis WBC 21.78 K/uL, moderate neutrophilia 18.67 K/uL, mild basophilia 0.25 K/uL, mild thrombocytosis 546 K/uL EPOC- Marked hypocalcemia 0.62 mmol/L (prev 4.3), Crea 0.39, rest wnl PCV 39% 12/24 PM bloodwork- ALB 1.4, PCV 45%, TS 3.2%

**ULTRASONOGRAPHIC EXAMINATION OF THE ABDOMEN**

**Urinary System**

The **urinary bladder**, trigone, and pelvic urethra presented normal thicknesses and normal tone. The ureters were not visible which is normal. No uroliths or sediment were visualized and anechoic urine was present. No evidence of inflammatory or neoplastic changes were noted. Ureteral papillae were normal.

The **kidneys** revealed largely normal size and structure, corticomedullary definition and ratio (cortex 1/3 of medulla) were essentially maintained with some age-related loss of curvilinear patterns regarding the capsule and C/M junction. The cortices presented largely uniform texture with some increased echogenicity expected for his age patient. Medullary structure differed distinctly from that of the cortex. Mineralization was present in the kidneys. The left kidney measured 2.5 cm. Slight pyelectasia was noted in the left kidney.

**Adrenal Glands**

The left **adrenal gland** was visualized and recognized as having normal shape, size, position and echogenicity for this breed. The phrenic vasculature, glandular echogenicity and detail were unremarkable. Capsule, cortex, and medullary definition were normal for this age patient. The left adrenal gland measured 0.4 cm.

The region of the **right adrenal gland** was unremarkable.

**Spleen**

The **spleen** in this patient was uniform, yet volume contracted. Hydration status should be assessed.

**Liver**

The **liver** was mildly subnormal in size with coarse architecture and increased portal markings. A history of hepatitis is likely. The gallbladder and common bile duct were unremarkable. The portal vein was visualized and appeared to be of normal size. No overt portosystemic shunting.



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**Gastrointestinal**

The **gastrointestinal tract** revealed diffuse, hyperechoic fogging or overlay throughout the small intestine as well as areas of mucosal striations and speckling. This striation + fogging effect appeared to exclusively affect the mucosal layer with the submucosa, muscularis and serosa left in-tact. Reactive mesentery was present associated with the serosa indicative of active inflammation. This is most consistent with protein losing enteropathy/lymphangiectasia. Full thickness biopsies or endoscopy guided biopsies would be ideal to confirm. No obstructive disease or obvious suspicion of neoplasia. A minor amount of chyme was noted in the stomach. The gastric wall was edematous.

**Pancreas**

The base and limbs of the **pancreas** were observed to be largely isoechoic to surrounding omental fat. Some mild parenchymal remodeling, however, with mild deviation from curvilinear normalcy was observed. Pancreatic duct and capsular irregularities were present consistent with age related changes. If pain upon imaging (+ Murphy sign) was present or if the patient is focally painful in subxyphoid palpation then low-grade smoldering chronic pancreatitis should be suspected.

**Free Abdomen/Other**

A mild amount of **ascites** and some pleural effusion noted through the diaphragm.

**ULTRASONOGRAPHIC FINDINGS**

- Mucosal fogging/gastritis pattern
- Microhepatica

**INTERPRETATION OF THE FINDINGS & FURTHER RECOMMENDATIONS**

Assessment for proteinuria warranted, if not already performed. If no significant proteinuria is present, then the albumin loss is likely from the GI tract owing to lymphangiectasia. Bile acid profile warranted given the hepatic presentation. Chest radiographs warranted to assess level of pleural effusion and other causes of pleural effusion as well as hypoalbuminemia.

**PLE Therapy**

Part or all of this protocol may be considered based on your clinical impression of the patient:

**OBJECTIVE: keep albumin levels > 2 g/dl, avoid thromboembolism and cavitory effusions, monitor concurrent PLN (Wheaton Terrier PLE/PLN) and liver disease:**

**Plasma** 10 mL / kilogram IV over 4 hours

Or **Human albumin** 2 ml/kg/h over 10 hours. Total daily volume 20.l/kg/day

**And Colloids/Hetastarch**

10 to 20 mL per kilogram per day and dogs

10 to 15 mL per kilogram per day cats

(Can bolus first 1/3 of dose over 15 minutes)

& maintain on LRS maintenance otherwise.

**Metronidazole** (10-20 mg/kg po bid)

**Famotidine** 1 mg/kg lv 1m po dc Sid /bid

**Sucralfate** 0.5-1 g po tid dogs, 0.5 g bid cats in slurry **Or Misoprostol** 1-5 ug/kg po tid



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**Diet:** Highly digestible high quality protein, low fiber, low fat diet (< 15% of dry matter). Hydrolyzed protein or novel protein. Purina HA or Royal Canine HP or similar.

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**Prednisone** or prednisolone 2 mg/kg bid x 3-5 days then 2 mg/kg sid. **Chlorambucil** in refractive severe IBD/alimentary lymphoma cases (monitor cbc for rare bone marrow suppression) 4 mg/m<sup>2</sup> Q 24-48 hours.

**BREED**

Yorshire Terrier

**Cobalamine** (B12) 250-1500 ug/dog weekly x 6 weeks.

**Calcium** supplementation if necessary.

**Aspirin** 0.5-1 mg/kg/day or **Clopidrel** (Plavix) 1-5 mg/kg/day.

**SEX**

Spayed Female

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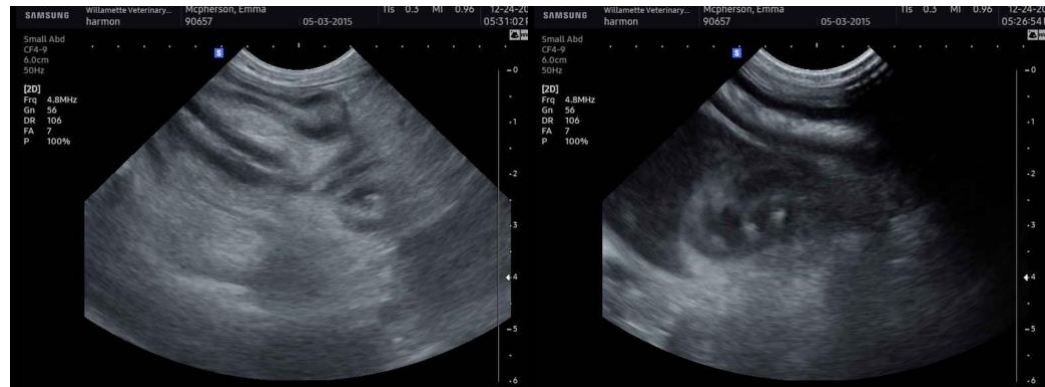
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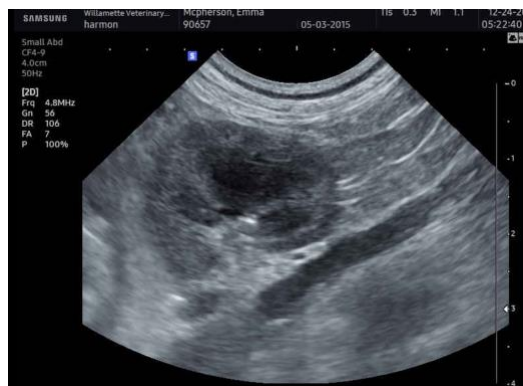
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The information and recommendations provided are based on the images presented by the referring veterinarian. No evaluation can be communicated regarding pathology that was not visible in the image/video clips provided.

Thank you for this referral. If the clinical or image interpretation does not parallel your findings or if I can be of any further assistance please contact me.

**Eric Lindquist**, DMV, DABVP, Cert. IVUSS, CEO of SonoPath.com  
[info@SonoPath.com](mailto:info@SonoPath.com)

**Protein-Losing Nephropathy (PLN)**

<http://www.sonopath.com/PLN>

**Description:** Protein-losing nephropathy (PLN) is a common form of renal disease that typically affects dogs in middle age; it occurs less commonly in cats. Glomerular causes of renal protein loss encompass two broad categories: glomerulonephritis (GN) and amyloidosis. (The causes of GN in human medicine are more specifically differentiated based on a combination of histopathology, immunofluorescence, and electron microscopy findings.) Membranoproliferative glomerulonephritis is the most common cause of GN in dogs and is associated with infectious disease with secondary immune complex deposition as well as Lyme disease. Membranous nephropathy is the second most common cause of GN in dogs and the most common cause in cats. It occurs due to primary immune complex deposition on the urinary side of the basement membrane of the glomerulus, resulting in the leakage of albumin. Amyloidosis is caused by the deposition of amyloid A proteins in a  $\beta$ -pleated sheet configuration in the glomeruli. It is a familial disease in the Shar Pei, but occurs as a reactive disease in other canine breeds. It is also inheritable in the Abyssinian cat, but the amyloidosis occurs in the medulla and is therefore not a protein-losing condition in this breed.

Glomerular lesions can be associated with:

- Infectious diseases:
  - Protozoan: *Babesia*, *Hepatozoon*, and *Leishmania*.
  - Bacterial: *Borrelia*, *Bartonella*, *Brucella*, *Ehrlichia*, *Mycoplasma*, pyometra, pyoderma, endocarditis, and pyelonephritis.



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- Viral: FeLV, FIV, and FIP.
- Fungal
- Helminthic: *Dirofilaria*.

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- Non-infectious inflammatory diseases: pancreatitis, chronic dermatitis, inflammatory bowel disease, periodontal disease, polyarthritis, and systemic lupus erythematosus (SLE).
- Neoplasia: lymphoma, leukemia, and mast cell disease.
- Familial conditions in the soft-coated Wheaten Terrier, Shar Pei, Beagle, Cocker Spaniel, and Bernese mountain dog.
- Idiopathic conditions.

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Post-glomerular causes, such as hemorrhage and inflammation, also contribute to urine protein quantification.

*Proteinuria Classifications:* Patients can be divided into three tiers, depending on their clinical characteristics:

**AGE**

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Tier 1A: persistent subclinical proteinuria

Tier 1B: persistent proteinuria with hypertension

**WEIGHT**

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Tier 2A: proteinuria and hypoalbuminemia

Tier 2B: proteinuria, hypoalbuminemia, and hypertension

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Tier 3A: proteinuria and azotemia

Tier 3B: proteinuria, azotemia, and hypertension

Tier 3C: proteinuria, azotemia, hypertension, and hypoalbuminemia

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**Diagnostics:** Traditionally, urine protein loss has been detected either through a qualitative test, such as a urine dipstick, or with a semi-quantitative test, such as a urine protein-creatinine (UPC) ratio. When the latter is greater than 0.5, it is considered abnormal. False positive results can occur due to contamination of urine with red blood cells, white blood cells, and bacterial protein. Thus, one must use a urine sample with inactive sediment and a negative culture for measurement purposes. A 24-hour urine protein quantification is more accurate but technically more difficult to obtain, as it requires hospitalization and 24-hour urinary catheterization with a closed collection system. Pooling urine samples can be considered in cases where urine protein loss is stable. One must obtain three different urine samples, combine 1 ml from each sample to submit for a UPC test, and ensure that inactive sediments are present in all the samples. There should be a high degree of correlation between the UPC on the pooled sample and the mean of the three samples measured independently. Research has not yet demonstrated the accuracy of pooled samples for urine samples with high protein loss (i.e., in cases where the UPC is > 8).



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Further diagnostic tests will depend on the tier classification. Once proteinuria is documented repeatedly, additional tests can be considered to assess for potential underlying causes, and, further to that, possible sources of antigen stimulation. Depending on presentation, tests may include:

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- CBC and biochemical profile
- Urine culture and sensitivity
- 4DX
- Blood pressure measurement
- Thoracic and abdominal radiographs
- Spinal radiographs to assess for discospondylitis
- Abdominal ultrasound to assess for evidence of underlying infection or neoplasia
- Echocardiogram to assess for vegetative endocarditis and possible effects of hypertension
- Screen for Cushing's disease, especially if hypertensive (LDDST or ACTH stimulation)
- ANA
- Expanded tick or infectious disease screen
- Renal biopsy to differentiate among specific causes of PLN

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Renal biopsy should be considered if proteinuria is severe (UPC > 3.5) and hypoalbuminemia and/or hypertension have been documented. Renal biopsy is an invasive procedure and should be considered only to determine if there is an underlying disease process that would benefit from specific therapy. If the patient is debilitated, severely azotemic, or has uncontrolled hypertension or coagulation abnormalities, then the risk of the procedure and anesthesia may be too great and should not be pursued.

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Tissue samples should be submitted for a combination of light microscopy (in formalin; use with special stains), immunofluorescence (in Michel's solution or frozen), and electron microscopy (in formalin with glutaraldehyde). It is imperative to request special media before obtaining the biopsy. Samples can be obtained via ultrasound guidance, laparotomy, or laparoscopy, but cortical samples must be divided so that they can be placed in the three different media. One must ensure that the pre-surgical clotting profile and platelet count are both normal. Patients should undergo pre-biopsy and post-biopsy diuresis.

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**Treatment:** The main goals of therapy are to i) reduce proteinuria (i.e., UPC < 1.0); ii) prevent a thrombotic event; iii) manage hypertension; and iv) replace fluid deficits. Fluid therapy should be approached cautiously, especially in patients with nephrotic syndrome. Standard therapy for PLN includes a low-protein diet, which in itself will reduce proteinuria, and the administration of an angiotensin-converting enzyme (ACE) inhibitor, such as enalapril (0.5 mg/kg PO BID) or benazepril (0.5 mg/kg PO Q24hr). Newer proposed therapeutic protocols include increasing the ACE inhibitor



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dose slowly while monitoring BUN and creatinine carefully. The dose can be raised to 1 mg/kg PO BID if needed, provided creatinine has not increased more than 30% from the baseline level.

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Another class of drugs currently being used is angiotensin receptor blockers, such as Losartan (the dose in azotemic dogs is 0.125-0.25 mg/kg/day PO Q12-24hr and 0.5-1.0 mg/kg/day in non-azotemic patients). This can be combined with an ACE inhibitor, but it is important to monitor BUN, creatinine, and potassium levels. Spironolactone has been used in people in combination with the other two classes of drugs to further modify the renin-angiotensin-aldosterone system (RAAS) (1-2 mg/kg PO BID); however, the effect of using all three drug classes in dogs has not yet been fully investigated. All of these medications are potassium sparing; thus, monitoring for hyperkalemia is important.

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Hypertension is managed with amlodipine (0.1-0.2 mg/kg PO Q12-24hr) when an ACE inhibitor is insufficient to control blood pressure. Supplementing with an anti-thrombotic agent, such as aspirin (1 mg/kg PO Q24hr), may be considered in advanced cases, especially once the patient is hypoalbuminemic. Omega-3 fatty acids can be given (0.25-0.5 g/day), but are typically increased in standard kidney diets.

**WEIGHT**

5.1 Lbs.

The most recent controversy in the management of glomerular diseases is the use of immunosuppressive medications. Because it is possible to arrive at a more definitive diagnosis in human patients, the use of immunosuppressive agents can be useful in the management of the disease, specifically when the disease is immune-mediated in its pathogenesis, such as SLE, membranous nephropathy, and minimal change disease glomerulonephritis. The procurement of a renal biopsy is being advocated in dogs so that practitioners can identify the population of patients that may benefit most from immunosuppressive therapy. Presently, there is no evidence-based medicine to suggest that immunosuppressive therapy should definitely be incorporated into a daily protocol for canine patients; however, it could be beneficial in some cases and may even result in remission. Further investigation is warranted. Trials are currently being conducted in patients with Lyme nephritis that are treated with immunosuppressive agents in addition to standard antibiotic therapy. The IRIS Treatment of Canine Glomerular Disease Study Group has suggested the trial use of immunosuppressive therapy in severe, persistent, or progressive PLN, even without a biopsy diagnosis in specific cases that are unresponsive to standard therapy (i.e., nephrotic syndrome, progressively azotemic, hypoalbuminemic patients). One can also consider administering the following drugs: pulse steroid therapy, mycophenolate, cyclophosphamide, azathioprine, and chlorambucil. One should monitor blood work, UPC ratio, and blood pressure weekly for 2 weeks, then biweekly for 6 weeks, then monthly. If there is further deterioration, immunosuppressive therapy should be discontinued.

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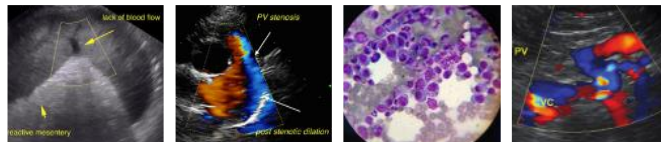
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**References:**



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Goldstein R and Polzin D. Treatment of canine glomerular disease: report of the IRIS treatment of canine glomerular disease study group. Proceedings from the American College of Veterinary Internal Medicine, Denver, CO, June 15-18, 2011.

#### SPECIES

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Grauer GF, Greco DS, Getzy DM, et al. Effects of enalapril versus placebo as a treatment for canine idiopathic glomerulonephritis. *J Vet Intern Med* 2000;14:526-33.

#### BREED

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Less GE, Cianciolo RE, and Clubb FJ. Renal biopsy and pathologic evaluation of glomerular disease. *Top Companion Anim Med* 2011;26(3):143-53.

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LeVine DA, Zhang D, Vaden SL. The use of pooled vs. serial urine samples to measure urine protein:creatinine ratios. *Vet Clin Pathol* 2010;39(1):53-56.

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Nability MB, Boggess MM, Kashtan CE, et al. Day-to-day variability in the urine protein:creatinine ratio in female dogs with stable glomerular proteinuria caused by x-linked hereditary nephropathy. *J Vet Intern Med* 2007;21:425-30.

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Vaden SL. Glomerular Disease. *Top Companion Anim Med* 2011;26(3):128-34.

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Vaden SL. Glomerular Diseases. In: Ettinger SJ and Feldman EC, eds. *Textbook of Veterinary Internal Medicine, 7<sup>th</sup> Ed.* St Louis, MI: Saunders Elsevier; 2010;2021-36.

#### HOSPITAL NAME

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Vaden SL. Microalbuminuria: What is it and how do I interpret it. Proceedings from the American College of Veterinary Internal Medicine, Charlotte, NC, June 4-7, 2003.

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Vaden SL et al. Urinary tract inflammation has a variable effect in urine albumin concentration. *J Vet Intern Med* 2002;16:378 (abstract).

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